

**Labs for course #412**  
**Analyzing Microarray Data using the mAdb System**  
**July 15-16, 2008 1:00pm- 4:00pm**

- First, look at the questions on the bottom of each page. Write down the answers while going through the steps on the page.
- Keep the browser NOT maximized so multiple windows can be distinguished.

# Lab 1. Copying a Training Dataset

Goal: To copy a dataset into user's temporary area and to inspect dataset features.

4

[Copy](#) Small, Round Blue Cell Tumors (SRBCTs) data containing 88 Arrays with 2308 Features to your temporary area.  
*Classification and diagnostic prediction of cancers using gene expression profiling and artificial neural networks.*  
Khan J, Wei JS, Ringer M, Saal LH, Ladanyi M, Westermann F, Berthold F, Schwab M, Antonescu CR, Peterson C, Meltzer PS, *Nature Medicine* Vol 7, Num 6, 601-673 (2001)

[Copy](#) Subset of NEJM data containing 60 Arrays with 1626 Features to your temporary area. Includes Feature Property Filters.

1. Open a web browser and type the URL for the mAdb home page, **for training class: <http://madb-training.cit.nih.gov>** - use login on name tent and password on board. Others can use <http://madb.nci.nih.gov> (NIAID users <http://madb.niaid.nih.gov>) and log in with your mAdb account.

2. Click the **mAdb Gateway** link to access mAdb Gateway Web page

3. On the mAdb Gateway Web page, click the link **Access Training/Public Dataset** on the bottom of the page. A page for copying three training datasets will be presented.

4. You can choose to work with either "Small, Round Blue Cell Tumors (SRBCT) dataset" or "NEJM Dataset". Click link **Copy** to copy the dataset into your temporary area.

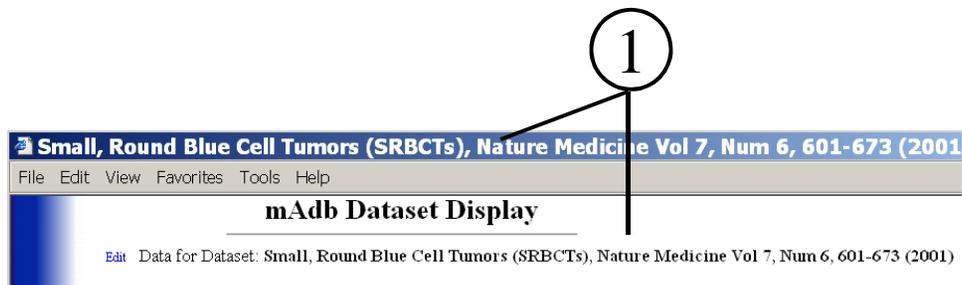
5. After copying the data, you will see the temporary dataset area. Click link **Open** on the selected dataset line. A mAdb Dataset Display page will be displayed.

Temporary Datasets		Created	Containing	Need Help?	
			Arrays	Genes	
<a href="#">Edit</a>	Small, Round Blue Cell Tumors (SRBCTs), Nature Medicine Vol ...	Aug 26 6:00:00pm	88	2308	<a href="#">Open</a> <a href="#">Refresh</a>
<a href="#">Edit</a>	NEJM - 3 Classes	Aug 26 5:23:18pm	60	1629	<a href="#">Open</a> <a href="#">Refresh</a>

5

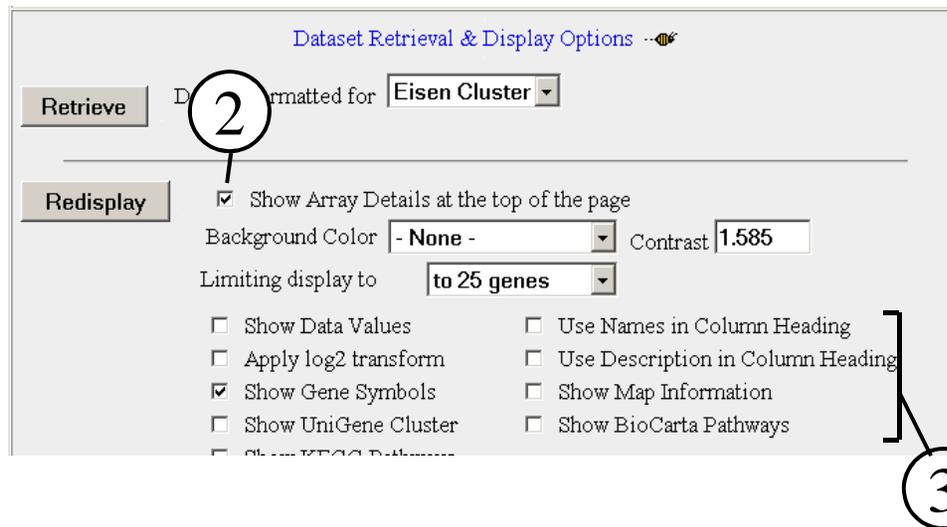
### Questions:

1. How many genes and how many arrays do you have in your dataset?



1. On the mAdb Dataset Display page, review the title bar, or the dataset description on top of the page. This tells you which dataset you are displaying.

2. In the **Dataset Retrieval & Display Options** panel, check the **Show Array Details at the top of the page** option. Then click **Redisplay** button. The names and short descriptions of arrays in the dataset will be displayed on the top of the page. Look for naming conventions of the array and then answer the question below. This information will be used in the next lab.



After reviewing the array details, it is recommended to uncheck the **Show Array Details at the top of the page** option. Click **Redisplay** to hide the array details on the top of the page.

3. Check or uncheck other display options of interest, and click **Redisplay** button to display or hide the relevant information. Uncheck **Show Data Values** and set **Background Color** to None will make it easier to view other annotations.

### Questions:

1. How many experiment groups can you identify in this dataset by their naming conventions? Write down the naming conventions for each group.

## Lab 2. Assigning Group Labels

Goal: To partition arrays into groups according to experiment design by assigning group labels.

The screenshot shows a web interface for data analysis. At the top, there is a section titled "Filtering/Grouping/Analysis Tools". Below this title, there are two main sections: "Choose a Tool" and "Choose a Viewer".

In the "Choose a Tool" section, a dropdown menu is set to "Filter/Group by Array Properties" (callout 1) and a "Proceed" button is visible (callout 2). Below this is the "Choose a Viewer" section, where a dropdown menu is set to "MDS: MultiDimensional Scaling" and a "View" button is visible.

The main part of the interface contains five groups (Group A through Group E). Each group has a dropdown menu for "Array Name" and another dropdown menu for "Begins with" (or "Contains" for Group E). Text input fields are provided for each group to specify the matching patterns. For Group A, the "Begins with" field contains "EWS" (callout 3). For Group B, it contains "BL". For Group C, it contains "NB". For Group D, it contains "RMS". For Group E, the "Contains" field is empty.

Below the group settings, there is a text input field for "Subset Label" containing "My Grouped Dataset" (callout 4). At the bottom left, there is a "Submit" button (callout 5) and a "Cancel" button at the bottom right.

Additional text below the group settings reads: "Expand the number of possible Group Designations to 10, 15, 20 or 26 groups."

In the Filtering/Grouping/Analysis section, choose the **Filter/Group by Array Properties Tool**

2. Click on **Proceed**

A new page will be displayed with options for assigning arrays into groups by the naming convention of **Array Name** or **Short Description**.

3. For the SRBC dataset, use EWS, BL, NB, RMS as matching patterns. Select **Array Name** and **Begins with** from the drop down list for each group. Samples with name beginning with "Test" are excluded from the grouped subset.

For the NEJM dataset, use GCB, ABC, and Type as matching patterns. Select **Array Name** and **Begins with** from the drop down list for each group.

4. The grouped results are stored as a new subset. Enter an appropriate label for this subset.

5. Click on **Submit**. There is no "Waiting" page, the new grouped subset will be directly displayed when the Group/Filtering process is completed.

## Lab 2. Assigning Group Labels

## mAdb Dataset Display

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[Edit](#) Data for Subset: **My Grouped Dataset**

from Dataset: **Small, Round Blue Cell Tumors (SRBCTs), Nature Medicine Vol 7, Num 6, 601-673 (2001)**

1

Filter/Group by Array Property

88 arrays and 2308 genes in the original dataset

63 arrays and 2308 genes in the output dataset.

Filter/Group by Array Property:

Group A: Array/Set Name Begins with 'ews'

Group B: Array/Set Name Begins with 'bl'

Group C: Array/Set Name Begins with 'nb'

Group D: Array/Set Name Begins with 'rms'

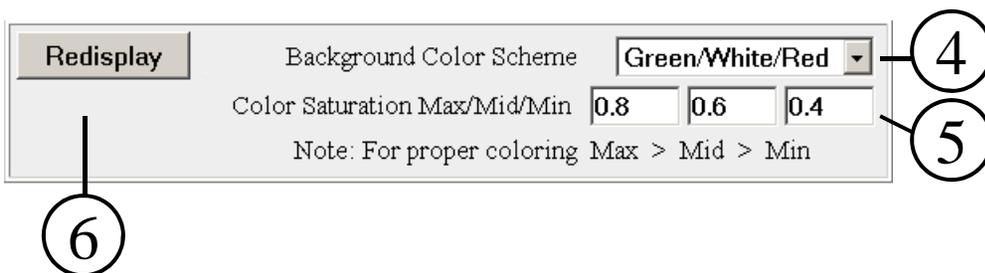
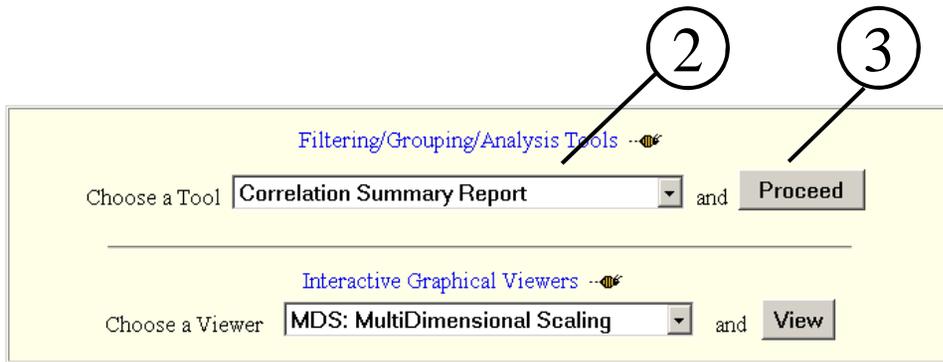
Examine the grouped subset through the dataset description and history on top of the Dataset Display page.

### Questions:

1. How many arrays are filtered out in the grouped dataset?
2. What are they? Hint use “Array Order Designation/Filtering Tool”.
3. How many arrays do you have in each group? Write down the group designations for each tumor type.

# Lab 3. Generating a Correlation Summary Report

Goal: To study the correlation of expression data among samples in the dataset.



- Verify that the current dataset is My Grouped Dataset through title bar or dataset description. (See Lab 1, Dataset display section for details)
- **2.** In the Filtering/Grouping/Analysis section, choose the **Correlation Summary Report** Tool. (You may have to scroll down the Tool dropdown list to find it on the bottom.)
- **3.** Click on **Proceed**.
- mAdb Correlation Report page will be displayed with a table of correlation results.
- **4.** Change the **Background Color Scheme** to **Green/White/Red**.
- **5.** Inspect the values of the correlation tables and set the values for **Color Saturation**. For SRBCT dataset, use 0.8, 0.6, 0.4. For NEJM-3 class dataset use 0.3, 0.0, -0.3.
- **6.** Click on **Redisplay** button. Correlation table will be colored according to the correlations.

## Lab 3. Generating a Correlation Summary Report

1. The image shows part of the correlation table. The color pattern uses green for good correlations and red for poor correlations.
2. Each correlation number represent a pair-wise correlation calculation between 2 samples. It can be clicked to display a scatter plot between the 2 samples. Click on a larger number to display the scatter plot for 2 correlated samples.
3. Click a small number to display a scatter plot for 2 poorly correlated samples .

B	B	B	B	B	B	B	B	C	C	C	C	C	C	C	C	C	C	C	C
#24	#25	#26	#27	#28	#29	#30	#31	#32	#33	#34	#35	#36	#37	#38	#39	#40	#41	#42	#43
#24 B	0.841	0.812	0.793	0.707	0.682	0.712	0.719	0.500	0.500	0.518	0.661	0.652	0.599	0.607	0.618	0.599	0.634	0.604	0.615
	#25 B	0.846	0.834	0.759	0.728	0.759	0.781	0.555	0.554	0.568	0.668	0.674	0.602	0.654	0.698	0.640	0.654	0.654	0.651
		#26 B	0.896	0.751	0.708	0.763	0.778	0.521	0.536	0.569	0.624	0.617	0.538	0.618	0.643	0.593	0.623	0.623	0.639
			#27 B	0.725	0.669	0.742	0.755	0.521	0.555	0.587	0.619	0.628	0.544	0.592	0.655	0.600	0.629	0.665	0.656
				#28 B	0.862	0.897	0.855	0.669	0.674	0.693	0.646	0.601	0.590	0.614	0.536	0.593	0.619	0.636	0.612
					#29 B	0.904	0.832	0.700	0.634	0.651	0.574	0.582	0.535	0.604	0.533	0.590	0.593	0.574	0.536
						#30 B	0.876	0.686	0.619	0.646	0.581	0.564	0.515	0.606	0.523	0.587	0.592	0.586	0.563
							#31 B	0.710	0.644	0.666	0.574	0.559	0.509	0.631	0.571	0.591	0.588	0.587	0.579
								#32 C	0.654	0.711	0.642	0.649	0.516	0.785	0.683	0.699	0.671	0.668	0.675
									#33 C	0.826	0.622	0.615	0.803	0.606	0.572	0.610	0.670	0.688	0.628
										#34 C	0.677	0.664	0.662	0.677	0.663	0.667	0.716	0.745	0.711
											#35 C	0.836	0.776	0.804	0.749	0.752	0.835	0.837	0.840
												#36 C	0.708	0.748	0.760	0.776	0.780	0.818	0.812
													#37 C	0.651	0.593	0.656	0.765	0.762	0.709
														#38 C	0.790	0.762	0.782	0.752	0.755
															#39 C	0.736	0.766	0.754	0.791
																#40 C	0.809	0.785	0.751
																	#41 C	0.873	0.777
																		#42 C	0.821
																			#43 C

1. Describe the general color pattern of the correlation table. Are correlation numbers within a group better (more green) than between groups (more red)?
2. How is the scatter plot of a good correlation different from a plot of a poor correlation?

## Lab 3. Generating a Correlation Summary Report

# Lab 4. Filtering Data

Goal: To pre-process a dataset for further analysis by filtering out genes with low variance or with many missing values.

Filtering/Grouping/Analysis Tools

Choose a Tool **Additional Filtering Options** and **Proceed**

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Interactive Graphical Viewers

Choose a Viewer **MDS: MultiDimensional Scaling** and **View**

Missing Value Filters

Genes: Require values in  $\geq$   % of Arrays

Arrays: Require values in  $\geq$   % of Genes

Gene Filters

Ratio  $\geq$   in  $\geq$   Arrays

Apply Symmetrically

Ratio  $\geq$   in  $\geq$   Arrays OR

Ratio  $\leq$   in  $\geq$   Arrays

Average Ratio  $\geq$

Apply Symmetrically

Max (Ratio) / Min (Ratio)  $\geq$

Variance (Gene Vector) percentile  $\geq$   %

Subset Label:

**Filter** **Cancel**

1. Use back button on web browser to return to previous Dataset Display page. Verify that the current dataset is My Grouped Dataset. (See Lab 1, Dataset display section for details).
2. In the Filtering/Grouping/Analysis section, choose the **Additional Filtering Options** Tool.
3. Click on **Proceed**
4. Data Filtering Options page will be displayed with options for Missing Value Filters and Gene Filters. Be careful to check the "checkboxes" along putting in values in step 4-9.
5. Select the check box for **Genes: Require values in  $\geq$**
6. Set the value to **95% of arrays**.
7. Select the check box for **Variance (Gene Vector) percentile**
8. Set the value  $\geq$  **90%**
9. The filtered results are stored as a new data subset. Enter an appropriate Label for this subset.
10. Click on **Filter**. Filtering will be performed and the results stored as a new subset. There is no "Waiting" page, the new subset will be directly displayed when the Filtering process is completed.

## Lab 4. Filtering Data

## mAdb Dataset Display

[Edit](#) Data for Subset: **Gene**>=95%, **Variance** >=90%  
from Dataset: **Small, Round Blue Cell Tumors (SRBCTs), Nature Medicine Vol 7, Num 6, 601-673 (2001)**

The filter input data set contained 63 arrays and 2308 genes.  
The filtered output data set contains 63 arrays and 230 genes.  
No genes excluded for being present in less than 95% (60) arrays.  
2078 genes excluded where variance is in the lowest 90 percentile (Variance<1.60).

View the complete [History](#).

1

2

1. Review the subset history on top of the Dataset Display page for the filtering.

2. Click link **History**, a new window will popup with the full dataset history. Review the text.

3. Click **output Dataset** will lead you to the filtered dataset. Close the new window and return to the previous window.

This dataset was constructed from the supplemental data posted at

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Thu Oct 9 17:57:21 EDT 2003

Filter/Group by Array Property  
88 arrays and 2308 genes in the [original dataset](#) dataset  
63 arrays and 2308 genes in the output dataset.

Filter/Group by Array Property:  
Group A: Array/Set Name Begins with 'ews'  
Group B: Array/Set Name Begins with 'bl'  
Group C: Array/Set Name Begins with 'nb'  
Group D: Array/Set Name Begins with 'rms'

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Fri Oct 10 10:35:40 EDT 2003

63 arrays, 2308 genes in the [input Dataset](#)  
230 Genes and 63 arrays passed filters  
No genes excluded for being present in less than 95% (60) arrays.  
2078 genes excluded where variance is in the lowest 90 percentile (Variance<1.60).

Link to the [output Dataset](#)

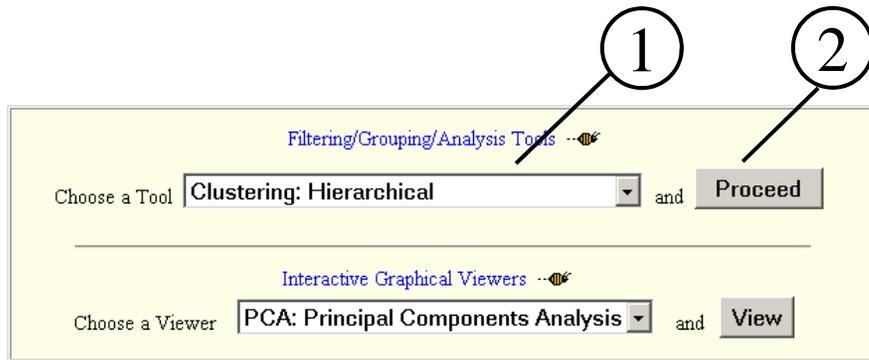
3

### Questions:

1. How many genes are filtered out by missing values? How many genes are filtered out by variance?

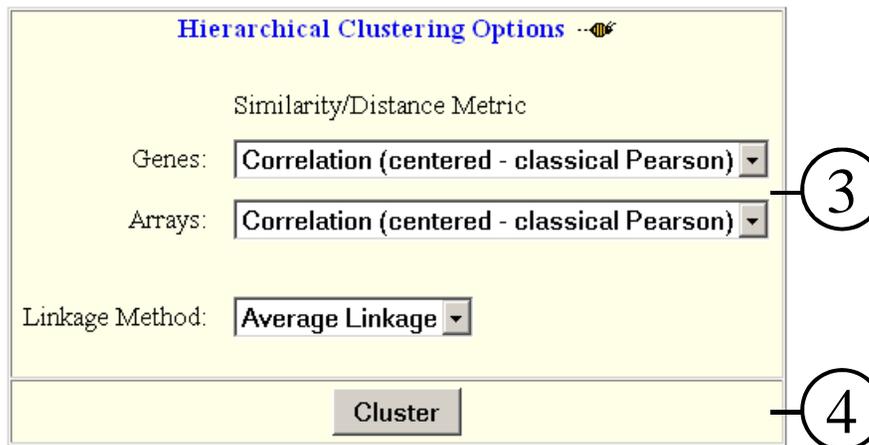
# Lab 5. Hierarchical Clustering

Goal: To cluster genes and/or arrays with the Hierarchical Clustering algorithm.



Verify that the current dataset is the filtered dataset.  
(Gene $\geq$ 95, Variance $\geq$ 90)

1. In the Filtering/Grouping/Analysis section, choose the **Clustering: Hierarchical** Tool.
2. Click on **Proceed**
3. A new page will be displayed with options for selecting the Similarity/Distance Metric.
4. Choose **Correlation (centered classical Pearson)** to cluster both Genes and Arrays.
5. Click on **Cluster** button.



## Lab 5. Hierarchical Clustering

A new page will be displayed for Hierarchical Clustering progress. When the analysis is done, a **View Clusters** button is displayed on top of the page.

1. Click the **View Clusters** button at the top of the page or the **Click to view result** link at the bottom.
2. A new page will be displayed with a thumbnail image of the clustering results

1

View Clusters

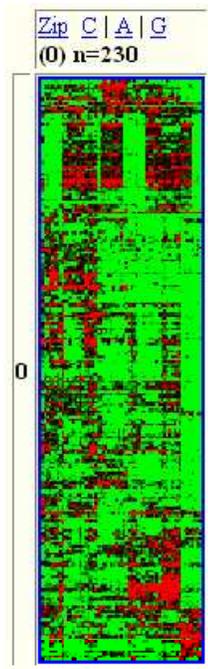
A **View button** should appear above when clustering is finished (a link will also appear at the bottom).

Clustering is performed using a derivative of the **Xcluster** program developed at Stanford University by [Gavin Sherlock](#), Head Microarray Informatics.

Initiating Hierarchical Clustering program...

```
Getting size of data...
Reading Data...
Done reading data...
Assigning Genes to Centroids: iteration 1
Assigning Genes to Centroids: iteration 2
Converged
Making correlations
0
Done Making Correlations
Clustering genes
Done clustering genes
Making correlations
0
Done Making Correlations
Clustering Experiments
10
20
30
40
50
60
Done Clustering Experiments
Outputting cdt file
Done outputting
Finished
```

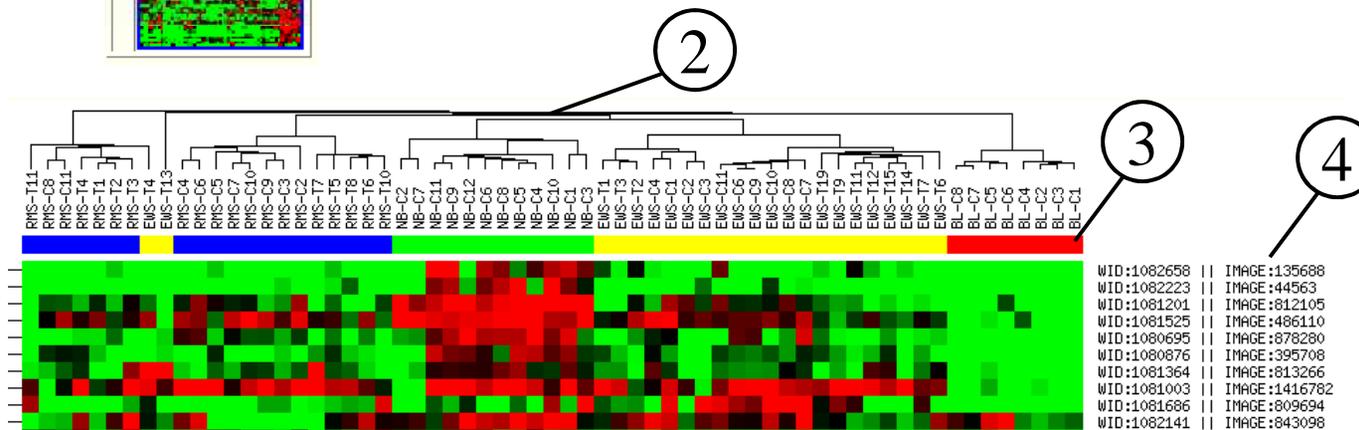
[Click to view result](#)



Click the thumbnail on the page.

1. A new browser window will open up to display an enlarged heatmap image, gene trees and array trees.
2. Check the array tree structure. Check the relationship among all the tumor groups.
3. The color bar indicate the grouping information of arrays. Identify the misclassified samples. Speculate possible explanations.
4. Click on the gene annotations on the right. A new window will open up with a feature report page.

Close the Feature Report window. Close the Heatmap display window. Return to the thumbnail image window.



### Questions:

1. How do the tumor samples cluster together? Can you find duplicate genes that cluster together on the heatmap?

## Lab 5. Hierarchical Clustering

## Lab 6. SOM Clustering

Goal: To partition genes into a 2-dimensional topology using the Self Organizing Map (SOM) algorithm and to observe genes with similar expression patterns.

The screenshot shows a web interface for SOM Clustering. It is divided into several sections:

- Filtering/Grouping/Analysis Tools**: Contains a dropdown menu for "Choose a Tool" set to "Clustering: SOM" and a "Proceed" button. A circled number 2 points to the dropdown, and a circled number 3 points to the "Proceed" button.
- Interactive Graphical Viewers**: Contains a dropdown menu for "Choose a Viewer" set to "MDS: MultiDimensional Scaling" and a "View" button.
- Data Adjustment Options**: Contains a dropdown menu for "Median Center Genes before Clustering" with a "NEW" label. A circled number 4 points to this dropdown.
- Self Organizing Maps Options**: Contains input fields for "Specify X dimension" (4), "Specify Y dimension" (3), and "Number of iterations" (100000). There is also a checkbox for "Initialize with Randomized Partition" which is unchecked. A circled number 5 points to the "Specify X dimension" field.
- SOM Elements Hierarchical Clustering Options**: Contains dropdown menus for "Genes" (Correlation (centered - classical Pearson)), "Arrays" (Not Clustered), and "Linkage Method" (Average Linkage). A circled number 6 points to the "Genes" dropdown.
- Cluster**: A button at the bottom of the interface. A circled number 7 points to this button.

Use the back button of the browser to return to the previous Dataset Display page. Verify that the current dataset is the right dataset. (Gene $\geq$ 95, Variance $\geq$ 90)

2. In the Filtering/Grouping/Analysis section, choose the **Clustering: SOM** Tool

3. Click on **Proceed**

A new page will be displayed with options for SOM.

4. Select **Median Center Genes before Clustering**

5. Set X dimension to be 4 and Y dimension to be 3, number of iterations to be 100000. Uncheck the checkbox for Initialize with Randomized Partition.

6. Set the Hierarchical Clustering Options within the SOM clusters. Select **Correlation (centered –classical Pearson)** Metric for Genes and **Not Clustered** for arrays.

7. Click on **Cluster** button.

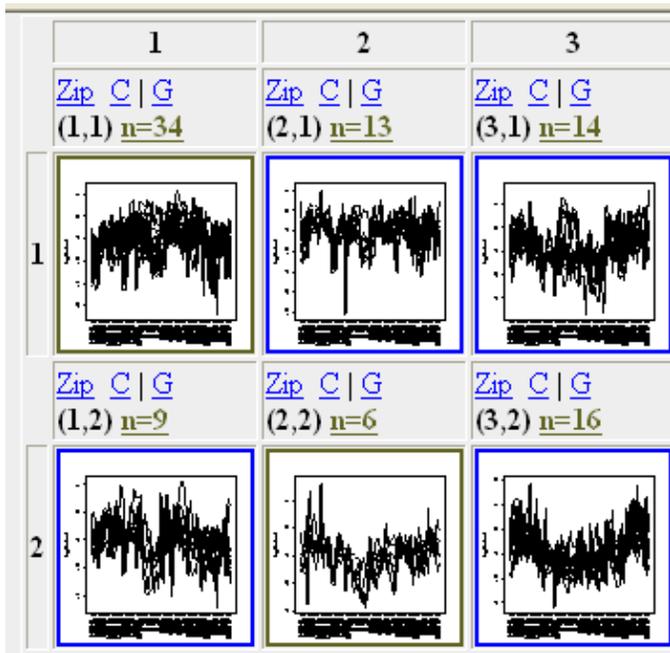
A new page will be displayed for SOM Clustering progress. When the analysis is done, a **View Clusters** button is displayed on top of the page.

8. Click the **View Clusters** button.

A new page will be displayed with a thumbnail image of the clustering results

## Lab 6. SOM Clustering





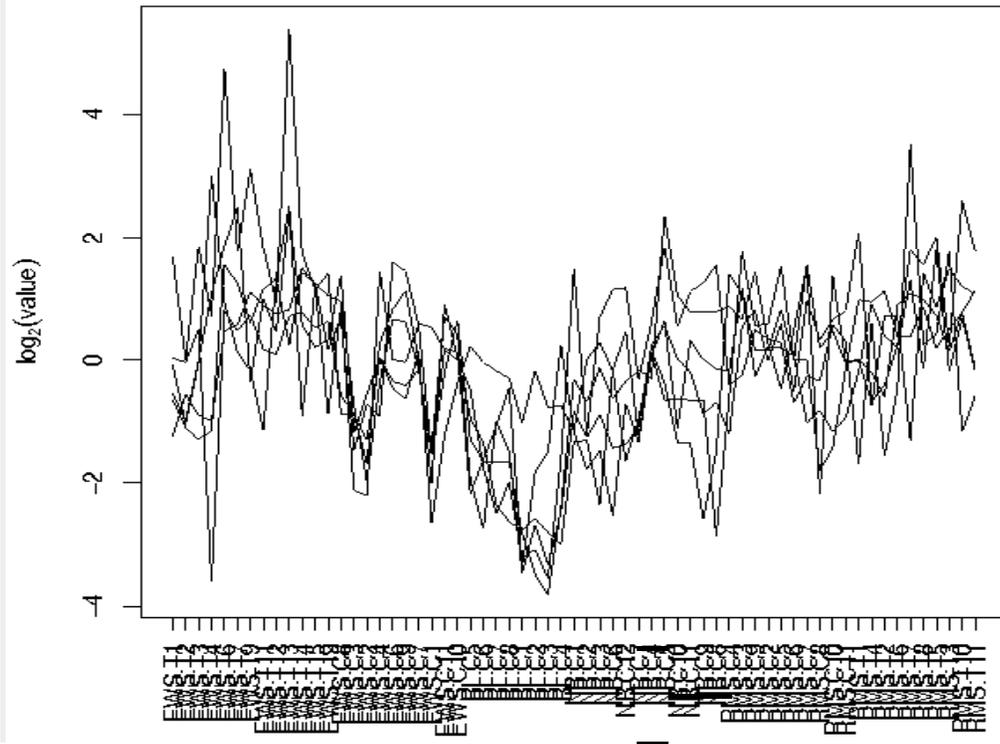
### Questions:

1. Do genes in the same partition show a similar expression profile? How are the expression profiles different among different partitions (2-D topology)?

1. Inspect the spatial relationship among the clusters.
2. Click a thumbnail image on the page.

A new browser window will open up to display an enlarged line plot image and gene tree of the clicked thumbnail image.

3. Can you interpret the graph?



# Lab 7. K-means Clustering (Optional)

Goal: To partition genes into  $K$  numbers of partitions using the K-means algorithm and observe genes with similar expression patterns.

2

Expand this Dataset.  
Access Datasets in your Temporary area.

Filtering/Grouping/Analysis Tools 

Choose a Tool  and

1. Use the back button of the browser to return to the previous Dataset Display page. Verify that the current dataset is the right dataset. (Gene $\geq$ 95, Variance $\geq$ 90)
2. Click link **Expand this Dataset** above the Filtering/Grouping/Analysis Tools section.

You will then be presented an expanded dataset selection page. You will find the dataset and all the subsets you saved from previous analysis.

3. Click link **Open** open My Grouped Dataset. A mAdb dataset display page will be presented to you. K-means clustering will be performed on the full grouped dataset to show its performance speed advantage.

3

Label	Origin	Created	Arrays	Genes	
<a href="#">Edit</a> Small, Round Blue Cell Tumors (SRBCTs), Nature Medicine Vol ...	Dataset	Aug 26 6:00:00pm	88	2308	<a href="#">Open</a>
<a href="#">Edit</a> My Grouped Dataset	Subset	Oct 09 5:57:20pm	63	2308	<a href="#">Open</a>
<a href="#">Edit</a> Gene $\geq$ 95%, Variance $\geq$ 90%	Subset	Oct 10 10:35:34 am	63	230	<a href="#">Open</a>

The screenshot shows a multi-step configuration interface for K-means clustering. It is divided into several sections:

- Filtering/Grouping/Analysis Tools:** Contains a dropdown menu set to "Clustering: Kmeans" (callout 1) and a "Proceed" button (callout 2).
- Interactive Graphical Viewers:** Contains a dropdown menu set to "MDS: MultiDimensional Scaling" and a "View" button.
- Data Adjustment Options:** Contains a dropdown menu set to "Median Center Genes before Clustering" (callout 3).
- Kmeans Clustering Options:** Contains two input fields: "Specify Number of Nodes" set to 12 (callout 4) and "Maximum Number of iterations" set to 100.
- Kmeans Nodes Hierarchical Clustering Options:** Contains two dropdown menus for "Similarity/Distance Metric": "Genes" is set to "Correlation (centered - classical Pearson)" (callout 5) and "Arrays" is set to "Not Clustered". Below these is a "Linkage Method" dropdown set to "Average Linkage".
- Cluster:** A large "Cluster" button at the bottom (callout 6).

1. In the Filtering/Grouping/Analysis section, choose the **Clustering: Kmeans Tool**.

2. Click on **Proceed**.

A new page will be displayed with options for Kmeans Clustering.

3. Select **Median Center Genes before Clustering**

4. Specify **Number of Nodes** to be 12. Set **Maximum Number of iterations** to be 100.

5. Set the Hierarchical Clustering Options within Kmeans Nodes. Select **Correlation (centered –classical Pearson)** for Genes and **Not Clustered** for arrays.

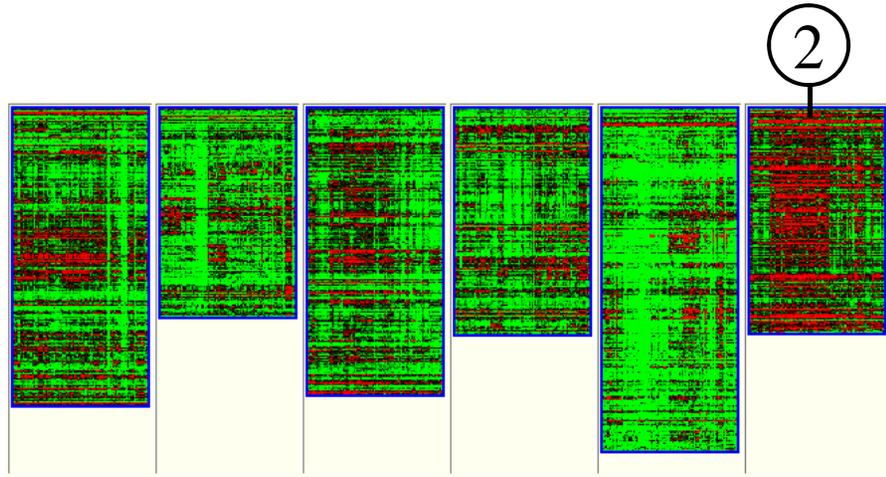
6. Click on **Cluster** button.

A new page will be displayed for Kmeans Clustering progress. When the analysis is done, a **View Clusters** button is displayed on top of the page.

7. Click the **View Clusters** button.

A new page will be displayed with a thumbnail image of the clustering results.

## Lab 7. K-means Clustering



1. Inspect the thumbnail images for the expression patterns within the clusters (Only 6 out of 12 clusters are displayed here).
2. Click thumbnails of interest on the page.

A new browser window will open up to display an enlarged heatmap image and gene tree (not shown here) of the clicked thumbnail image.

3. Close the Heatmap display window. Return to the thumbnail image window.

### Questions:

1. Are the expression profiles different among different partitions?
2. Can you interpret the expression profile of a given gene?

## Lab 7. K-means Clustering

# Lab 8. PCA

Goal: To explore the data structure of the dataset using Principal Component Analysis (PCA).

Filtering/Grouping/Analysis Tools

Choose a Tool  and

---

Interactive Graphical Viewers

Choose a Viewer  and

2 3

4 5

Perform PCA on:  Arrays  Genes

Dispersion Matrix:  Correlation  Covariance

Note: Imputing of missing values is not yet available.  
Genes with missing values are disgarded from the PCA calculations.

6

PCA was performed on 63 arrays and 230 genes.  
No genes contained a missing value.

Proceed to the

7

1. Verify that the current dataset is the filtered dataset.  
(Gene $\geq$ 95, Variance $\geq$ 90)
2. In the **Interactive Graphical Viewers** section, choose the viewer, **PCA: Principal Components Analysis**.

3. Click on **View** button.

A new window, PCA Options, will be displayed with options for the PCA Analysis .

4. Select to perform PCA on **Arrays**.

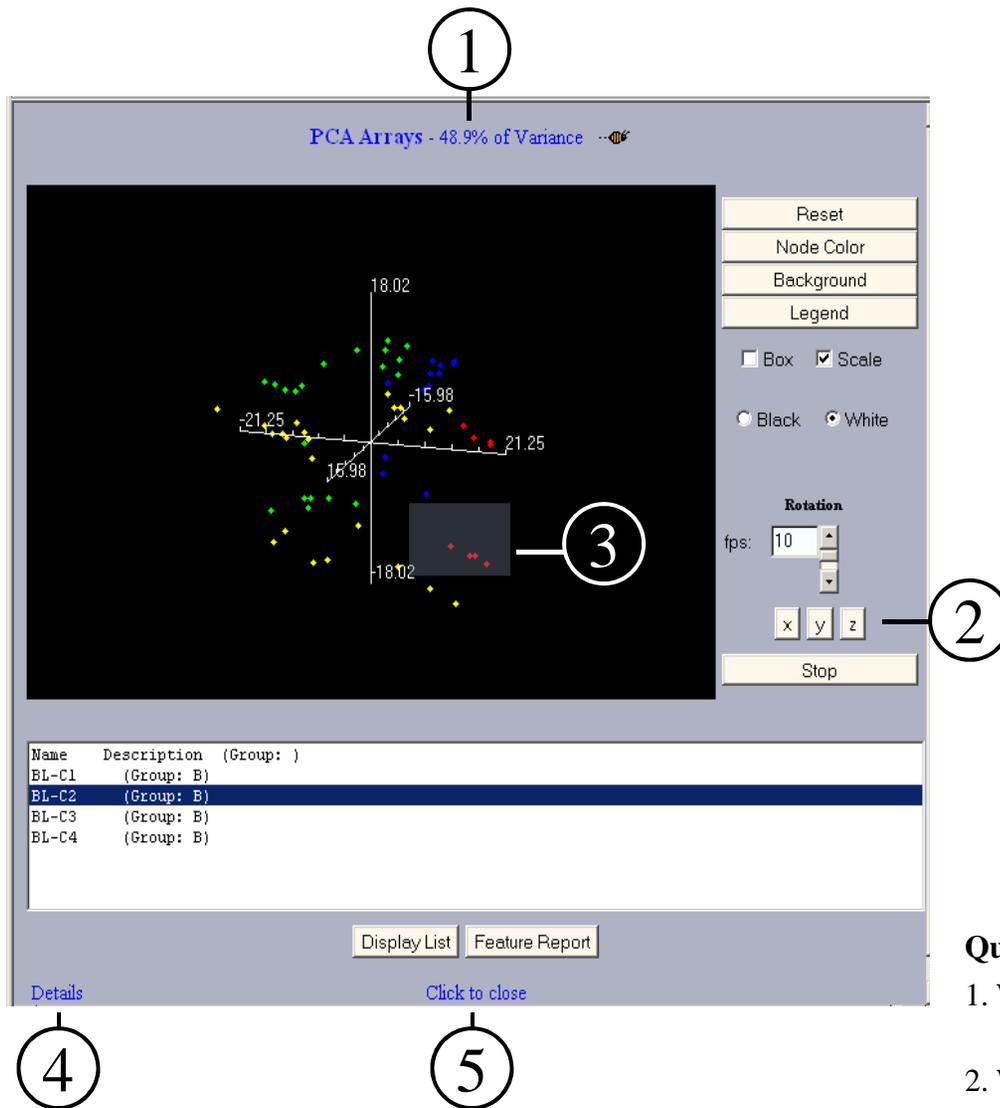
5. Select Dispersion Matrix of **Covariance**.

6. Click **Continue** button.

A new page, **Waiting for PCA**, will be displayed. When PCA analysis is done, a summary and a new button, **3D Viewer** will be displayed on the page.

7. Click **3D Viewer** button.

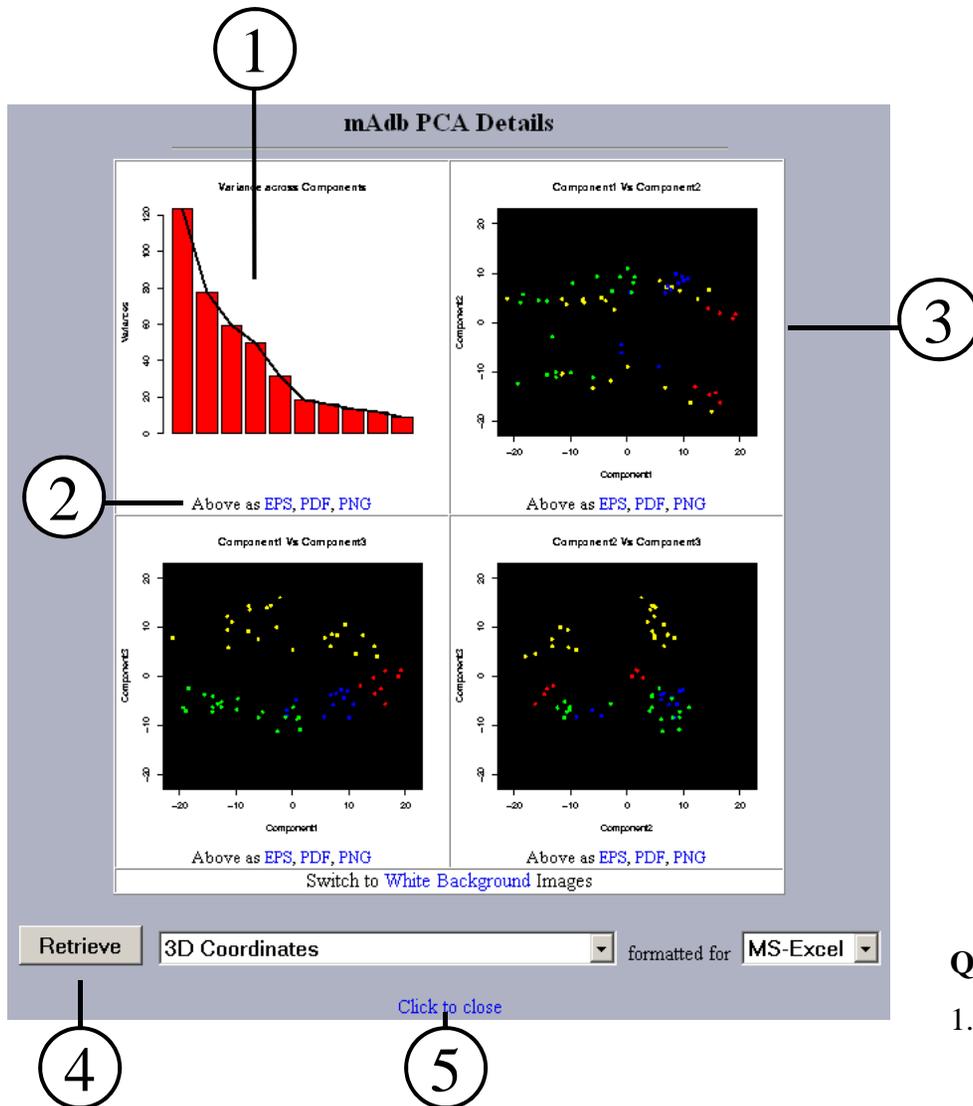
- **Questions:**
- 1. How many genes are used in PCA analysis?  
(Genes with missing values are not used in PCA)



1. Check the percentage of Variance represented in the 3D plot. Does it capture a large percentage of total variance?
2. Click the **X, Y and Z** buttons to rotate the 3-D plot. Look for clustering /separation of data. Click **Stop** button.
3. Click and Drag the mouse to highlight an area of the 3 D plot. Data points in the area will be displayed in the text area below the plot.
4. Click the link **Details** on the bottom of the 3D viewer. A new page, **PCA details**, will be displayed with 4 additional plots from PCA analysis. See next page for more description of the PCA details page.

### Questions:

1. What is the percentage of variance represented in the first three components?
2. What is the color-coding for each group of samples? Can you see a separation of different groups in 3D plot?



1. The Scree Plot displays the Variance for individual components. Click on the plot will display a new page with an enlarged image.
2. The PDF (Portable Document Format) or PNG (Portable Network Graphics) links under each figure can be used to display or save a larger image of the figure. You can also save a larger image as Encapsulated PostScript using the EPS link
3. The other three plots shown are 2-D plots for each combination of the first 3 components.
4. The **Retrieve** button will retrieve the data back to your local computer. Several options are available. We do not need to retrieve data for this Lab.
5. Click the link **Click to Close** to close the viewer. This will allow you to go back to the starting dataset display page.

### Questions:

1. In the scree plot, identify where the slope of variance flattens out (the scree point).

**Labs for course #412**  
**Analyzing Microarray Data using the mAdb System**  
**July 15-16, 2008 1:00pm- 4:00pm**

- First, look at the questions on the bottom of each page. Write down the answers while going through the steps on the page.
- Keep the browser NOT maximized so multiple windows can be distinguished.

Day 2

# Lab 9. Performing an ANOVA analysis

Goal: To identify differentially expressed genes using class comparison statistical tools.

Choose one or more Projects, select a Tool and Continue or access previously extracted data located in **training01**'s: [Temporary](#) area

1

1. On the mAdb Gateway Page, Click on **Temporary** area to open a list of your Datasets stored in this area.

Temporary Datasets		Created	Containing		Need Help?			Gene Information
			Arrays	Genes				Refreshed
<a href="#">Edit</a>	Small, Round Blue Cell Tumors (SRBCTs), Nature Medicine Vol ...	Aug 26 6:00:00pm	88	2308	<a href="#">Open</a>	<a href="#">Expand (1)</a>	<a href="#">Refresh</a>	Aug 26 6:00:00pm
<a href="#">Edit</a>	NEJM - 3 Classes	Aug 26 5:23:18pm	60	1629	<a href="#">Open</a>	<a href="#">Expand (1)</a>	<a href="#">Refresh</a>	Aug 26 5:23:18pm

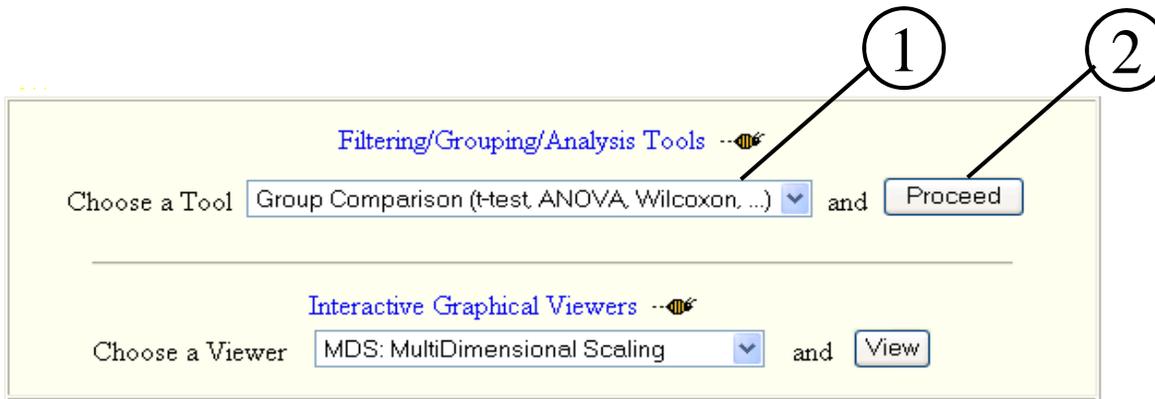
2

Label	Origin	Created	Containing		Need Help?	
			Arrays	Genes		
<a href="#">Edit</a>	Small, Round Blue Cell Tumors (SRBCTs), Nature Medicine Vol ...	Dataset Aug 26 6:00:00pm	88	2308	<a href="#">Open</a>	
<a href="#">Edit</a>	My Grouped Dataset	Subset Oct 16 2:12:33pm	63	2308	<a href="#">Open</a> <a href="#">History</a>	

3

2. Click on the **Expand** for the “Small Round Blue Cell Tumors (SRBCTs)...” (or, if you are using the other dataset, Expand for the “NEJM – 3 Classes”) to open the list of Subsets for this Dataset.
3. Click on the **Open** for the “My Grouped Dataset” subset.

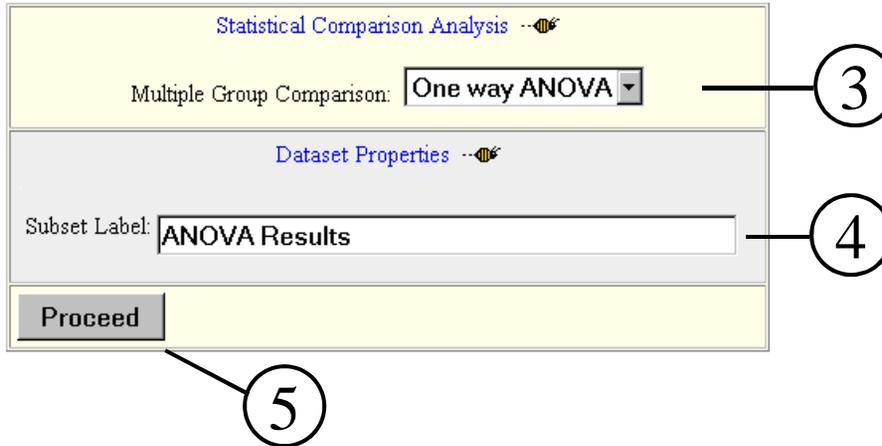
## Lab 9. Performing an ANOVA analysis



1. In the Filtering/Grouping/Analysis section, choose the **Group Comparison Tool**

2. Click on **Proceed**

A new page will be displayed with options for the statistical comparison analysis. Since this dataset has more than two groups, only the Multiple Group Comparison options for more than two groups will be available for selection.

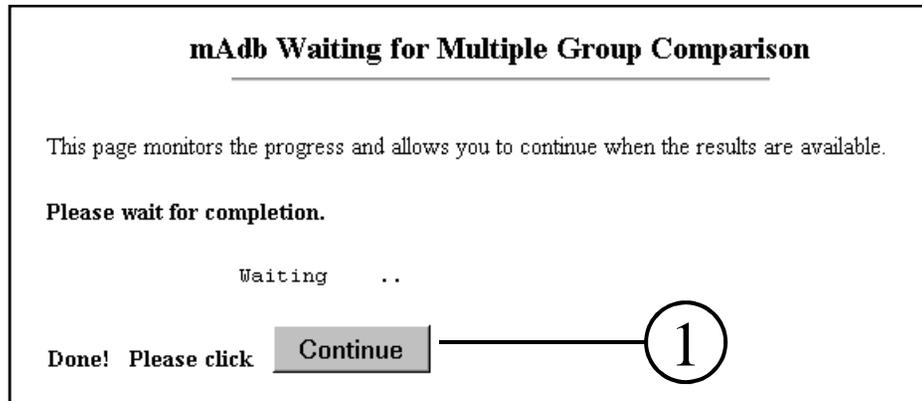


3. Select **One way ANOVA**

4. Analysis results are stored as a new subset. Enter an appropriate Label for this subset.

5. Click on **Proceed**.

## Lab 9. Performing an ANOVA analysis



A “Waiting” page is displayed while the analysis is being performed. When the analysis is completed, the continue button is displayed.

1. Click on **Continue**. A mAdb Dataset Display page, displaying the newly created subset which contains the ANOVA analysis results will appear.

The three columns, p-Value, Difference and Groups display results from this analysis. The p-Value is the One way ANOVA calculation. The Difference displays the largest difference between group means--this calculation is independent of the ANOVA calculation. The Groups identifies the two groups having this largest mean difference. The default order of the data is from smallest to largest p-Value.

Note that “Show Data Values” has been unchecked for the display shown here.

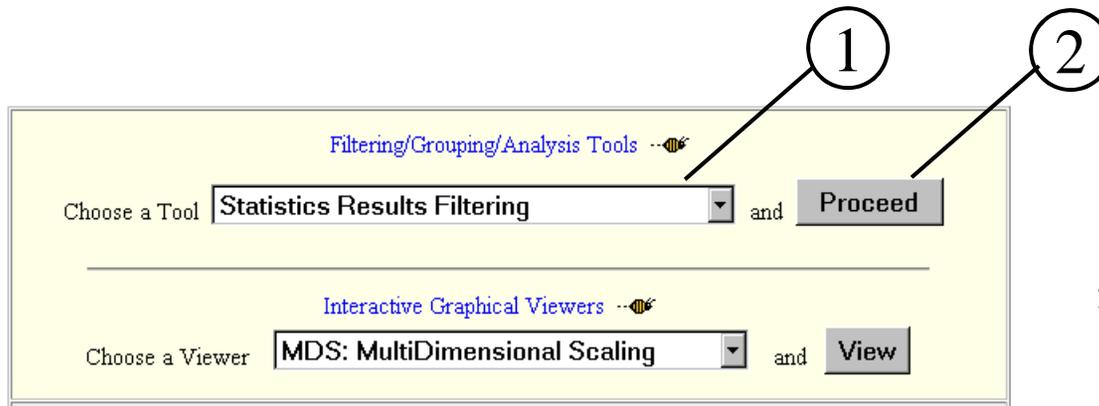
p-Value                       Difference  
 Groups

[Save a Feature Property List \(used with the Feature Properties Filtering tool\).](#)

Records 1 to 25 of 2308 total records displayed.

p-Value	Difference	Groups	Well ID	Feature ID	Gene	Description
9.6276e-22	4.11	A-B	1081848	<a href="#">IMAGE:770394</a>	FCGRT	Fc fragment of IgG, receptor, tra
3.488e-20	2.99	C-D	1082414	<a href="#">IMAGE:784224</a>	FGFR4	FGFR4=Fibroblast growth factor
2.5008e-19	3.59	A-B	1080705	<a href="#">IMAGE:377461</a>	CAV1	caveolin 1, caveolae protein, 22k
2.5733e-18	2.59	A-C	1082413	<a href="#">IMAGE:814260</a>	FVT1	fmt1=Follicular lymphoma variant
1.4459e-17	2.76	C-A	1081462	<a href="#">IMAGE:796258</a>	SGCA	sarcoglycan, alpha (50kDa dystro
5.7703e-17	2.89	A-B	1081004	<a href="#">IMAGE:1435862</a>	MIC2	antigen identified by monoclonal
8.728e-17	3.14	C-B	1081653	<a href="#">IMAGE:859359</a>	PIG3	quinone oxidoreductase homolog
1.3957e-16	3.95	D-A	1082509	<a href="#">IMAGE:295985</a>		clone IMAGE:4538214=FLJ2065
4.1114e-16	4.03	A-B	1080566	<a href="#">IMAGE:365826</a>	GAS1	Growth arrest-specific 1

## Lab 9. Performing an ANOVA analysis



1. In the Filtering/Grouping/Analysis section, choose the **Statistical Results Filtering** Tool

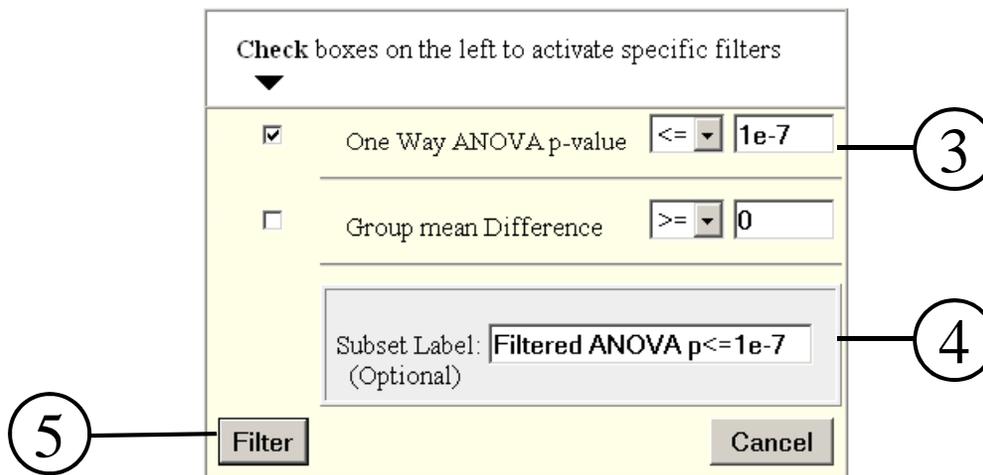
2. Click on **Proceed**

A new page will be displayed with the options for filtering the statistical results.

3. **Check** the box to the left of **One way ANOVA p-value**, select “<=“ and enter the p-value as **0.000001** or **1e-7**.

4. The filtered results will be stored as a new subset. Enter an appropriate Label for this subset.

5. Click on **Filter**. Filtering will be performed and the results stored as a new subset. There is no intermediate “Waiting” page, the new subset will be directly displayed when the Filtering process is completed.



### Questions:

1. How many genes are there in the filtered dataset?

In order to facilitate later comparison/filtering of these results with other results, we will save this result as a Feature Property List.

p-Value       Difference  
 Groups

Save a Feature Property List (used with the Feature Properties Filtering tool).

Records 1 to 25 of 388 total records displayed.

1

p-Value	Difference	Groups	Well ID	Feature ID	Gene	Description
9.6276e-22	4.11	A-B	1081848	IMAGE:770394	FCGRT	Fc fragment of IgG, receptor, tra
3.488e-20	2.99	C-D	1082414	IMAGE:784224	FGFR4	FGFR4=Fibroblast growth factor
2.5008e-19	3.59	A-B	1080705	IMAGE:377461	CAV1	caveolin 1, caveolae protein, 22k
2.5733e-18	2.59	A-C	1082413	IMAGE:814260	FVT1	fv1=Follicular lymphoma variant
1.4459e-17	2.76	C-A	1081462	IMAGE:796258	SGCA	sarcoglycan, alpha (50kDa dystro
5.7703e-17	2.89	A-B	1081004	IMAGE:1435862	MIC2	antigen identified by monoclonal
8.728e-17	3.14	C-B	1081653	IMAGE:859359	PIG3	quinone oxidoreductase homolog
1.3957e-16	3.95	D-A	1082509	IMAGE:295985		clone IMAGE:4538214=FLJ206:
4.1114e-16	4.03	A-B	1080566	IMAGE:365826	GAS1	Growth arrest-specific 1

2

Feature Property List

Save a List of: **mAdb Well IDs**

Store the List as: **Global (Available in all Datasets)**

List Label: **SRBCT ANOVA p<=1e-7**

Overwrite any existing list with the same label

Save

3

4

5

1. Click on Save a Feature Property List. A new page will be displayed with the options for the Saving a Feature Property List..
2. Select **mAdb Well IDs**
3. Select **Global (Available in all Datasets)**
4. Enter an appropriate label to identify this List
5. Click on **Save**

## Lab 9. Performing an ANOVA analysis

Successfully stored the list.

- Action: **Saved New global List**
- Type: **Well ID**
- Labeled: **SRBCT ANOVA  $p \leq 1e-7$**
- Containing: **141** unique, non empty elements

To return to the dataset/subset click

**Continue**

1

A page indicating that the List was successfully stored and summarizing information about the list will be displayed.

**1.** Click on **Continue**. This will return you back to the Data Display Page.

## Lab 9. Performing an ANOVA analysis

# Lab 10. Using SAM

Goal: To evaluate statistically significant genes and determine the False Discovery Rate (FDR).

Choose one or more Projects, select a Tool and Continue or access previously extracted data located in **training01**'s: [Temporary](#) area

1

1. On the mAddb Gateway Page, Click on **Temporary** area to open a list of your Datasets stored in this area.

Temporary Datasets	Created	Containing Arrays	Genes	Need Help?	Gene Information Refreshed
<a href="#">Edit</a> Small, Round Blue Cell Tumors (SRBCTs), Nature Medicine Vol ...	Aug 26 6:00:00pm	88	2308	<a href="#">Open</a> <a href="#">Expand (1)</a> <a href="#">Refresh</a>	Aug 26 6:00:00pm
<a href="#">Edit</a> NEJM - 3 Classes	Aug 26 5:23:18pm	60	1629	<a href="#">Open</a> <a href="#">Expand (1)</a> <a href="#">Refresh</a>	Aug 26 5:23:18pm

2

Label	Origin	Created	Containing Arrays	Genes	Need Help?
<a href="#">Edit</a> Small, Round Blue Cell Tumors (SRBCTs), Nature Medicine Vol ...	Dataset	Aug 26 6:00:00pm	88	2308	<a href="#">Open</a>
<a href="#">Edit</a> My Grouped Dataset	Subset	Oct 16 2:12:33pm	63	2308	<a href="#">Open</a> <a href="#">History</a>

3

2. Click on the **Expand** for the “Small Round Blue Cell Tumors (SRBCTs)...” (or, if you are using the other dataset, Expand for the “NEJM – 3 Classes”) to open the list of Subsets for this Dataset.
3. Click on the **Open** for the “My Grouped Dataset” subset.

## Lab 10. Using SAM

Filtering/Grouping/Analysis Tools

Choose a Tool **Filter/Group by Array Properties** and **Proceed**

---

Interactive Graphical Viewers

Choose a Viewer **MDS: MultiDimensional Scaling** and **View**

Group A **Array Name** **Begins with** BL

Group B **Array Name** **Begins with** NB

Group C **Array Name** **Contains**

Group D **Array Name** **Contains**

Group E **Array Name** **Contains**

Expand the number of possible Group Designations to 10, 15, 20 or 26 groups.

Subset Label: **Two groups for SAM - BL and NB only**

**Submit** **Cancel**

1. In the Filtering/Grouping/Analysis section, choose the **Filter/Group by Array Properties Tool**
2. Click on **Proceed**

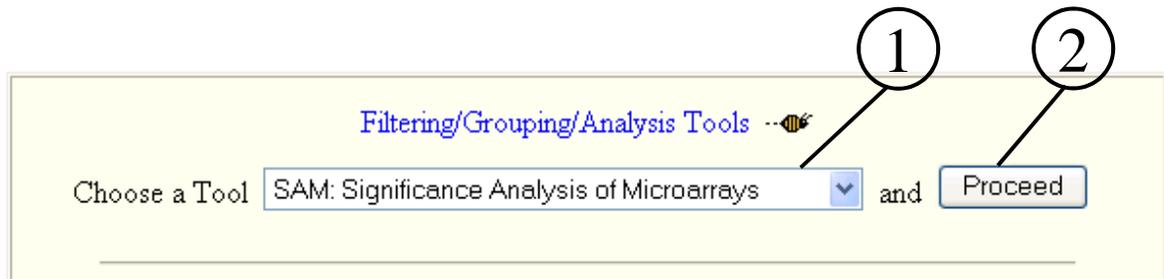
A new page will be displayed with options for assigning arrays into groups by the naming convention of **Array Name** or **Short Description**.

3. For the SRBC dataset, use BL and NB as matching patterns. Select **Array Name** and **Begins with** from the drop down list for each group.

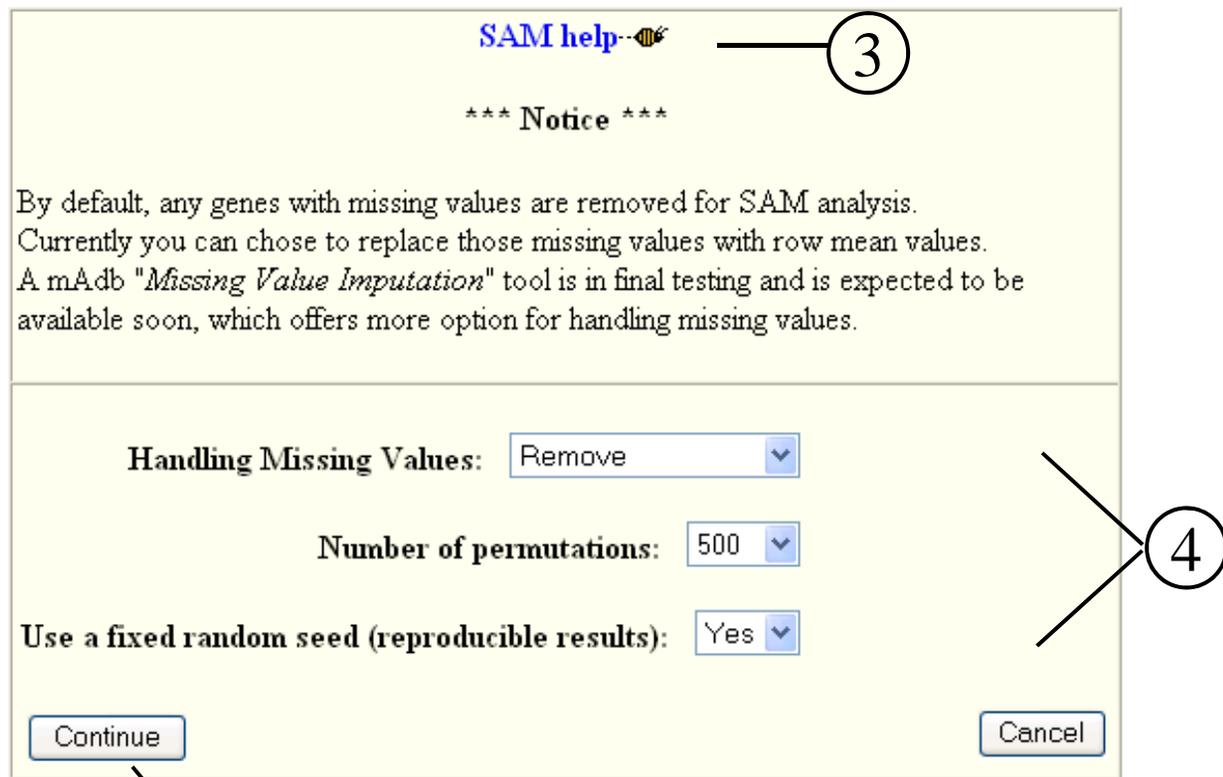
For the NEJM dataset, use GCB and ABC as matching patterns. Select **Array Name** and **Begins with** from the drop down list for each group.

4. The grouped results are stored as a new subset. Enter an appropriate label for this subset.
5. Click on **Submit**. There is no “Waiting” page, the new grouped subset will be directly displayed when the Group/Filtering process is completed.

## Lab 10. Using SAM



### mAdb SAM Options



1. In the Filtering/Grouping/Analysis section, choose the **SAM: Significance Analysis for Microarrays** Tool.
2. Click on **Proceed**.
3. Click on **SAM help**.
4. Select SAM options to **Remove** missing values, to perform **500** permutations, and set **Yes** for a fixed random seed.
5. Click on **Continue**.

## mAddb: Waiting for SAM

This page monitors the progress and allows you to continue when the results are available.

**Please wait for completion.**

Waiting .....

**SAM Step 1: FDR Calculations Completed!**

**SAM Analysis performed on 20 arrays and 2308 genes.  
No genes contained missing values.**

**Proceed to the**

SAM Step 2

1

SAM Analysis is initiated and a “waiting” page is displayed. When the Analysis is complete, an analysis summary and a button to continue to the next step appear on the page.

**1. Click on SAM Step2.**

### Questions:

1. How many genes contain missing values?

Clicking on a Delta value to create a new subset or on an image icon to generate the corresponding SAM plot;  
 or input a Delta value at the bottom and Click "Create Subset".

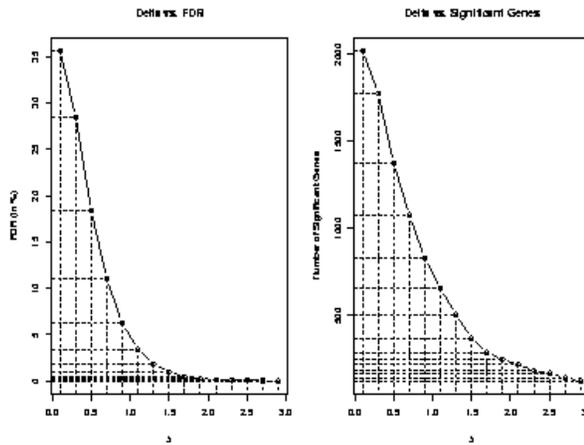
Delta	# of Sig. Genes	# of False Positives	FDR*
0.1	2020	719.87	0.3564
0.2	1946	646.87	0.3324
0.3	1776	505.57	0.2847
0.4	1605	376.30	0.2345
0.5	1374	252.76	0.1840
0.6	1194	171.35	0.1435
0.7	1077	118.13	0.1097
0.8	929	77.12	0.0830
0.9	829	51.30	0.0619
1.0	735	34.15	0.0465
1.1	654	22.35	0.0342
1.2	587	14.92	0.0254
1.3	504	9.16	0.0182
1.4	428	5.54	0.0129
1.5	370	3.53	0.0095
1.6	340	2.45	0.0072
1.7	287	1.40	0.0049
1.8	267	0.96	0.0036
1.9	245	0.64	0.0026
2.0	232	0.42	0.0018
2.1	218	0.28	0.0013
2.2	197	0.20	0.0010
2.3	182	0.13	0.0007
2.4	176	0.08	0.0005
2.5	169	0.07	0.0004
2.6	157	0.04	0.0003
2.7	142	0.03	0.0002
2.8	135	0.02	0.0002
2.9	125	0.02	0.0001

1

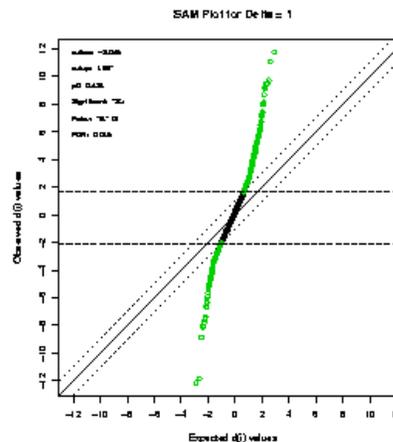
2

Create Subset

FDR: (# of False Positives)/(# of Sig. Genes)



Above as [EPS](#), [PDF](#), [PNG](#)



Above as [EPS](#), [PDF](#), [PNG](#)

The SAM results are displayed as a table and three graphs. The table shows the number of significant genes, the number of false genes and the false discovery rate (FDR) for each Delta. You can create a subset containing the genes corresponding to one of the models by either clicking on a Delta value or entering a Delta value in the text box and clicking the “Create Subset” button.

The top left graph plots the Delta vs. the FDR. The top right graph plots the Delta vs. the number of significant genes.

The lower graph plots the observed  $d(i)$  vs. expected  $d(i)$ , with a delta cutoff that generates the biggest FDR that is smaller than 0.05.

Click on an **image icon** with a low FDR (new window pops up.)

Click on a **Delta** with a low FDR.

**Questions:**

1. How many genes do you have for this Delta?
2. What is the FDR for the Delta?

- |   |   |
|---|---|
| <input type="checkbox"/> Show Data Values                 | <input type="checkbox"/> Use Names in Column Heading          |
| <input type="checkbox"/> Apply log2 transform             | <input type="checkbox"/> Use Description in Column Heading    |
| <input checked="" type="checkbox"/> Show Gene Symbols     | <input checked="" type="checkbox"/> Show Map Information      |
| <input checked="" type="checkbox"/> Show UniGene Cluster  | <input checked="" type="checkbox"/> Show BioCarta Pathways    |
| <input type="checkbox"/> Show KEGG Pathways               |   |
| <input type="checkbox"/> Show GO Tier 2 Component         | <input type="checkbox"/> Show GO Tier 3 Component             |
| <input type="checkbox"/> Show GO Tier 2 Function          | <input type="checkbox"/> Show GO Tier 3 Function              |
| <input type="checkbox"/> Show GO Tier 2 Process           | <input type="checkbox"/> Show GO Tier 3 Process               |
| <input checked="" type="checkbox"/> Show Gene Description | <input type="checkbox"/> Show GO Terms                        |
| <input type="checkbox"/> Show Average(Log2 Ratio)         | <input type="checkbox"/> Show Max(Log2 Ratio)-Min(Log2 Ratio) |
| <input type="checkbox"/> Show Variance                    |   |
| <input checked="" type="checkbox"/> d.value               | <input checked="" type="checkbox"/> Stand. Deviation          |
| <input checked="" type="checkbox"/> q.value               | <input checked="" type="checkbox"/> Fold Change               |

- A mAdb Dataset Display page, displaying the newly created SAM subset appears.
- Note that **Show Data Values** has been unchecked and the **Background Color** has been set to None for the display shown here.

Save a Feature Property List (used with the Feature Properties Filtering tool).

Records 1 to 25 of 370 total records displayed.

d.value	Stand. Deviation	q.value	Fold Change	Well ID	Feature ID	Map	UniGene	Gene
-12.1298	0.2684	0	0.0518	1081305	<a href="#">IMAGE:183337</a>	6p21.3	Hs.77522	HLA-DMA
-11.8486	0.3205	0	0.0384	1082374	<a href="#">IMAGE:840942</a>	6p21.3	Hs.814	HLA-DPB1
11.7632	0.2149	0	12.3195	1081310	<a href="#">IMAGE:563673</a>	5q31	Hs.74294	ALDH7A1
11.0799	0.2100	0	10.7428	1081326	<a href="#">IMAGE:784593</a>	2q23.3	Hs.6838	ARHE
9.7225	0.2372	0	9.1553	1081886	<a href="#">IMAGE:504791</a>	6p12.1	Hs.169907	GSTA4
9.5226	0.2314	0	8.3336	1082121	<a href="#">IMAGE:377048</a>	2q12-q34	Hs.121576	MYO1B
9.5000	0.3766	0	18.2186	1082060	<a href="#">IMAGE:629896</a>	5q13	Hs.103042	MAP1B
9.4193	0.3259	0	12.8741	1081201	<a href="#">IMAGE:812105</a>	1q21	Hs.75823	AF1Q
9.2278	0.2293	0	7.7811	1082481	<a href="#">IMAGE:204545</a>	2p13.1	Hs.8966	TEM8
9.1644	0.3089	0	14.6853	1080695	<a href="#">IMAGE:878280</a>	4p16.1-p15	Hs.155392	CRMP1
-8.8426	0.2167	0	0.1633	1081617	<a href="#">IMAGE:814526</a>	20q13.31	Hs.236361	RNPC1
8.6979	0.3444	0	11.1301	1081525	<a href="#">IMAGE:486110</a>	3q25.1-q25.2	Hs.91747	PFN2
8.1327	0.3580	0	11.4990	1082603	<a href="#">IMAGE:308231</a>	2q12-q34	Hs.121576	MYO1B

# Lab 11. Using PAM

Goal: To evaluate shrunken centroid prediction models and identify sets of genes that best classify sample types.

Choose one or more Projects, select a Tool and Continue or access previously extracted data located in **training01**'s: [Temporary](#) area

1

1. On the mAdb Gateway Page, Click on **Temporary** area to open a list of your Datasets stored in this area.

Temporary Datasets		Created	Containing		Need Help?			Gene Information
			Arrays	Genes			Refreshed	
<a href="#">Edit</a>	Small, Round Blue Cell Tumors (SRBCTs), Nature Medicine Vol ...	Aug 26 6:00:00pm	88	2308	<a href="#">Open</a>	<a href="#">Expand (1)</a>	<a href="#">Refresh</a>	Aug 26 6:00:00pm
<a href="#">Edit</a>	NEJM - 3 Classes	Aug 26 5:23:18pm	60	1629	<a href="#">Open</a>	<a href="#">Expand (1)</a>	<a href="#">Refresh</a>	Aug 26 5:23:18pm

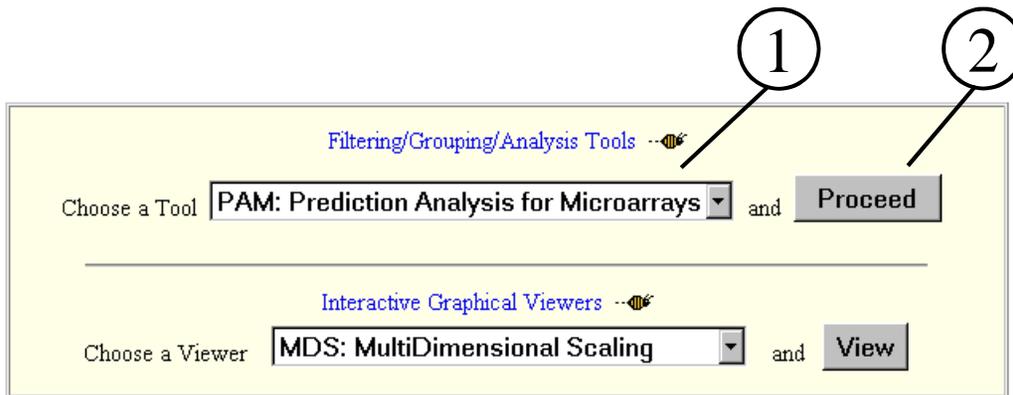
2

Label		Origin	Created	Containing		Need Help?	
				Arrays	Genes		
<a href="#">Edit</a>	Small, Round Blue Cell Tumors (SRBCTs), Nature Medicine Vol ...	Dataset	Aug 26 6:00:00pm	88	2308	<a href="#">Open</a>	
<a href="#">Edit</a>	My Grouped Dataset	Subset	Oct 16 2:12:33pm	63	2308	<a href="#">Open</a>	<a href="#">History</a>

3

2. Click on the **Expand** for the “Small Round Blue Cell Tumors (SRBCTs)...” (or, if you are using the other dataset, Expand for the “NEJM – 3 Classes”) to open the list of Subsets for this Dataset.
3. Click on the **Open** for the “My Grouped Dataset” subset.

## Lab 11. Using PAM

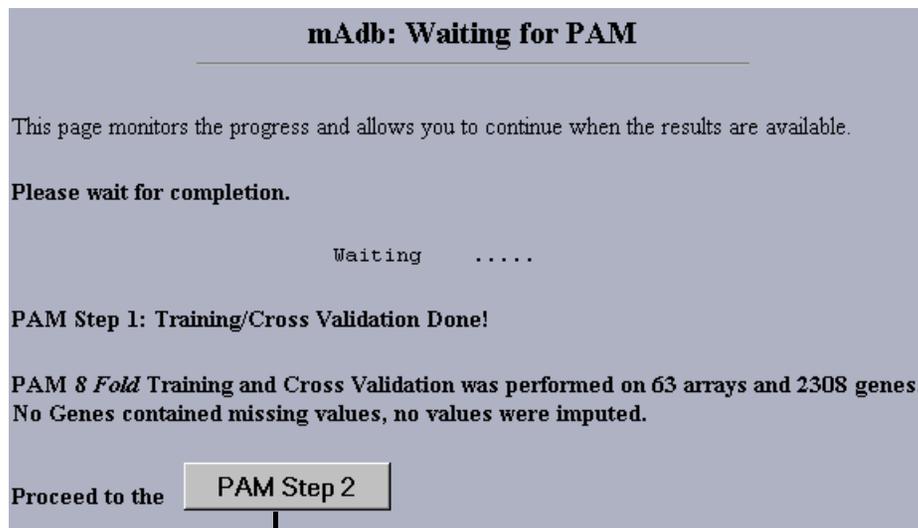


1. In the Filtering/Grouping/Analysis section, choose the **PAM: Prediction Analysis for Microarrays Tool**.

2. Click on **Proceed**

PAM Analysis is initiated and A “waiting” page is displayed. When the Analysis is complete, an analysis summary and a button to continue to the next step appears on the page.

3. Click on **PAM Step2**.



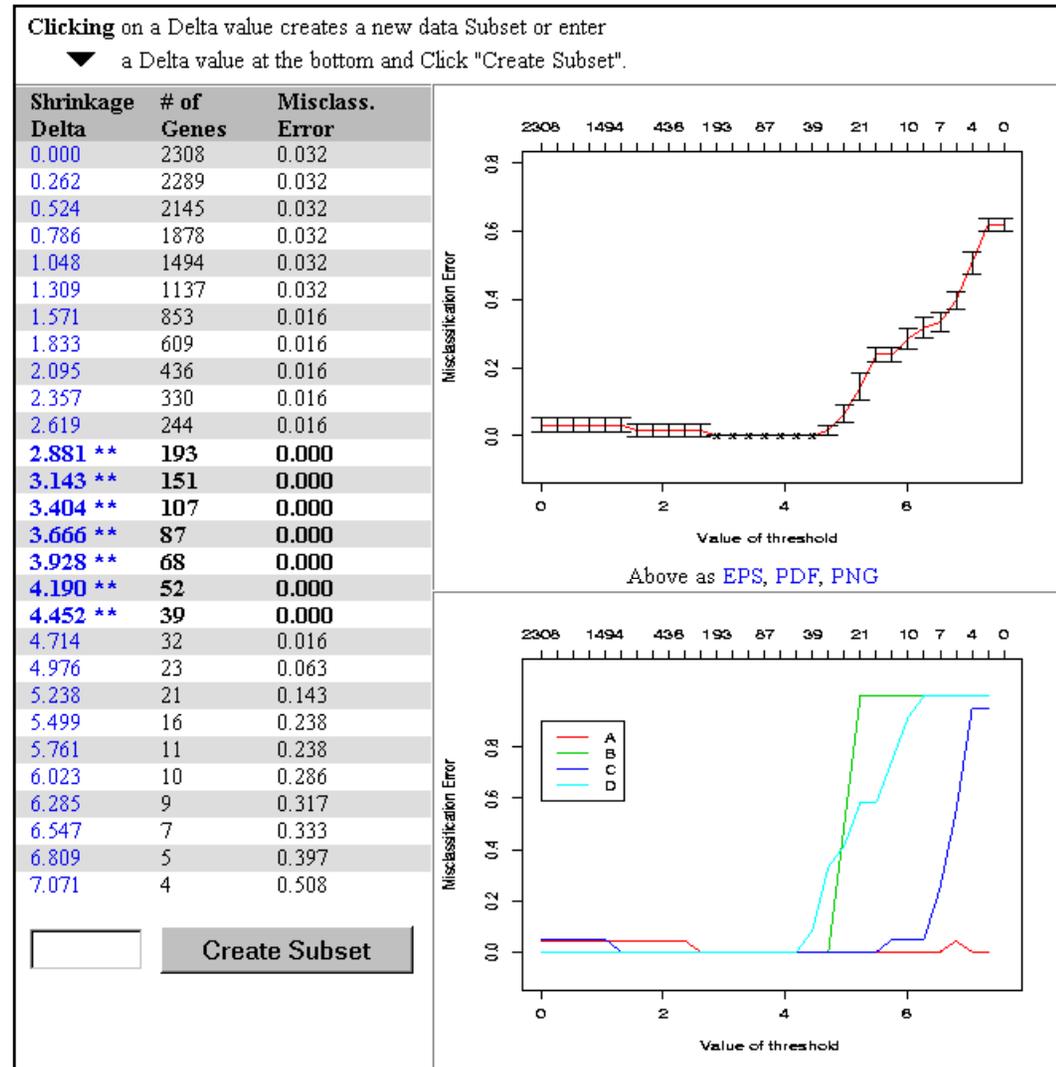
**Questions:**

1. How many fold of Training and Cross Validation was performed?
2. How many genes contain missing values? How many missing values are imputed for the dataset?

The PAM results are displayed as a table and two graphs. The table shows the Shrinkage Delta (\*\* indicates those having minimum misclassification error), number of genes in the model and the misclassification error based on the K-fold cross validation. You can create a subset containing the genes corresponding to one of the models by either clicking on a Shrinkage Delta value or entering a Delta value in the text box and clicking the “Create Subset” button.

The top graph plots the misclassification error (with error bars) versus the Shrinkage Delta (bottom axis) and the number of Genes (top axis).

The lower graph plots the misclassification error for each group versus the Shrinkage Delta (bottom axis) and the number of Genes (top axis).



## Lab 11. Using PAM

- Click on a Shrinkage Delta having a minimum misclassification error.

1

**Questions:**

- How many genes do you have in the model ?
- What is the Misclassification Error percentage for the model?

Clicking on a Delta value creates a new data Subset or enter a Delta value at the bottom and Click "Create Subset".

Shrinkage Delta	# of Genes	Misclass. Error
0.000	2308	0.032
0.262	2289	0.032
0.524	2145	0.032
0.786	1878	0.032
1.048	1494	0.032
1.309	1137	0.032
1.571	853	0.016
1.833	609	0.016
2.095	436	0.016
2.357	330	0.016
2.619	244	0.016
<b>2.881 **</b>	<b>193</b>	<b>0.000</b>
<b>3.143 **</b>	<b>151</b>	<b>0.000</b>
<b>3.404 **</b>	<b>107</b>	<b>0.000</b>
<b>3.666 **</b>	<b>87</b>	<b>0.000</b>
<b>3.928 **</b>	<b>68</b>	<b>0.000</b>
<b>4.190 **</b>	<b>52</b>	<b>0.000</b>
<b>4.452 **</b>	<b>39</b>	<b>0.000</b>
4.714	32	0.016
4.976	23	0.063
5.238	21	0.143
5.499	16	0.238
5.761	11	0.238
6.023	10	0.286
6.285	9	0.317
6.547	7	0.333
6.809	5	0.397
7.071	4	0.508

2308 1494 436 193 87 39 21 10 7 4 0

Misclassification Error

Value of threshold

Above as EPS, PDF, PNG

2308 1494 436 193 87 39 21 10 7 4 0

Misclassification Error

Value of threshold

— A  
— B  
— C  
— D

Create Subset

- A mAdb Dataset Display page, displaying the newly created PAM subset appears.
- The columns A Score, B Score, ... contain the shrunken differences for each group. Non zero values can be used to infer which group or groups a gene's expression value distinguishes.
- Note that **Show Data Values** has been unchecked and the **Background Color** has been set to None for the display shown here.

<input checked="" type="checkbox"/> A Score	<input checked="" type="checkbox"/> B Score
<input checked="" type="checkbox"/> C Score	<input checked="" type="checkbox"/> D Score

[Save](#) a Feature Property List (used with the Feature Properties Filtering tool).

Records 1 to 25 of 87 total records displayed.

A Score	B Score	C Score	D Score	Well ID	Feature ID	Gene	Description
0.6527	-0.1732	0.0000	0.0000	1081848	<a href="#">IMAGE:770394</a>	FCGRT	Fc fragment of IgG, receptor, transp
-0.0429	0.0000	0.6346	0.0000	1082414	<a href="#">IMAGE:784224</a>	FGFR4	FGFR4=Fibroblast growth factor rec
-0.1131	0.0000	0.6248	0.0000	1080646	<a href="#">IMAGE:296448</a>	IGF2	Insulin-like growth factor 2 (somato
0.0000	-0.6186	0.0000	0.0000	1082657	<a href="#">IMAGE:212542</a>		Homo sapiens mRNA; cDNA DKF
-0.5856	0.0000	0.0000	0.0000	1082509	<a href="#">IMAGE:295985</a>		clone IMAGE:4538214=FLJ20653
0.5773	0.0000	0.0000	0.0000	1080705	<a href="#">IMAGE:377461</a>	CAV1	caveolin 1, caveolae protein, 22kDa
0.0000	-0.5739	0.0000	0.0000	1082481	<a href="#">IMAGE:204545</a>	TEM8	tumor endothelial marker 8
0.0000	-0.5527	0.0000	0.0000	1081310	<a href="#">IMAGE:563673</a>	ALDH7A1	aldehyde dehydrogenase 7 family, nr
0.0000	0.0000	0.5420	0.0000	1080968	<a href="#">IMAGE:207274</a>		Homo sapiens cDNA: FLJ22066 fis

## Lab 11. Using PAM

In order to facilitate later comparison/filtering of these results with other results, we will save this result as a Feature Property List.

1. Click on Save a Feature Property List.

A new page will be displayed with the options for the Saving a Feature Property List..

A Score       B Score  
 C Score       D Score

Save a Feature Property List (used with the Feature Properties Filtering tool).

Records 1 to 25 of 87 total records displayed.

1

A Score	B Score	C Score	D Score	Well ID	Feature ID	Gene	Description
0.6527	-0.1732	0.0000	0.0000	1081848	<a href="#">IMAGE:770394</a>	FCGRT	Fc fragment of IgG, receptor, transp
-0.0429	0.0000	0.6346	0.0000	1082414	<a href="#">IMAGE:784224</a>	FGFR4	FGFR4=Fibroblast growth factor rec
-0.1131	0.0000	0.6248	0.0000	1080646	<a href="#">IMAGE:296448</a>	IGF2	Insulin-like growth factor 2 (somato
0.0000	-0.6186	0.0000	0.0000	1082657	<a href="#">IMAGE:212542</a>		Homo sapiens mRNA; cDNA DKF
-0.5856	0.0000	0.0000	0.0000	1082509	<a href="#">IMAGE:295985</a>		clone IMAGE:4538214=FLJ20653
0.5773	0.0000	0.0000	0.0000	1080705	<a href="#">IMAGE:377461</a>	CAV1	caveolin 1, caveolae protein, 22kDa
0.0000	-0.5739	0.0000	0.0000	1082481	<a href="#">IMAGE:204545</a>	TEM8	tumor endothelial marker 8
0.0000	-0.5527	0.0000	0.0000	1081310	<a href="#">IMAGE:563673</a>	ALDH7A1	aldehyde dehydrogenase 7 family, m
0.0000	0.0000	0.5420	0.0000	1080968	<a href="#">IMAGE:207274</a>		Homo sapiens cDNA: FLJ22066 fis

2

Feature Property List

Save a List of:

Store the List as:

List Label:

Overwrite any existing list with the same label

3

4

5

2. Select **mAdb Well IDS**
3. Select **Global (Available in all Datasets)**
4. Enter an appropriate label to identify this List
5. Click on **Save**

# Lab 12. Applying Hierarchical Clustering to the PAM Model

Goal: To use Hierarchical Clustering to explore a PAM Model.

Choose one or more Projects, select a Tool and Continue or access previously extracted data located in **training01**'s: [Temporary](#) area

1

1. On the mAdb Gateway Page, Click on **Temporary** area to open a list of your Datasets stored in this area.

Temporary Datasets		Created	Containing		Need Help?			Gene Information
			Arrays	Genes			Refreshed	
<a href="#">Edit</a>	Small, Round Blue Cell Tumors (SRBCTs), Nature Medicine Vol ...	Aug 26 6:00:00pm	88	2308	<a href="#">Open</a>	<a href="#">Expand (1)</a>	<a href="#">Refresh</a>	Aug 26 6:00:00pm
<a href="#">Edit</a>	NEJM - 3 Classes	Aug 26 5:23:18pm	60	1629	<a href="#">Open</a>	<a href="#">Expand (1)</a>	<a href="#">Refresh</a>	Aug 26 5:23:18pm

2

2. Click on the **Open** for the “Small Round Blue Cell Tumors (SRBCTs)...” (or, if you are using the other dataset, Expand for the “NEJM – 3 Classes”)

Filtering/Grouping/Analysis Tools

Choose a Tool **Feature Property Filtering Options** and **Proceed**

Interactive Graphical Viewers

Choose a Viewer **MDS: MultiDimensional Scaling** and **View**

1. In the Filtering/Grouping/Analysis section, choose the **Feature Property Filtering Options Tool**

2. Click on **Proceed**

A new page will be displayed with options for the Feature Property Filtering.

3. **Check, Include Only** where Well ID is in Feature List saved in previous Lab (SRBCT – 87 Gene PAM Model for SRBCT dataset ).

4. Enter an appropriate Label for the Subset.

5. Click on **Filter**.

Check boxes on the left to activate specific filters

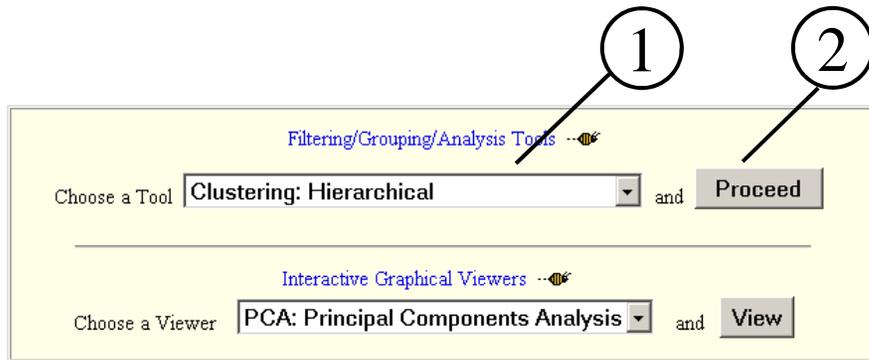
- Exclude** Designated Housekeeping Genes
- Include only** Designated Control Features
- Include only** where Well ID =
- Include only** where  =< Well ID <=
- Include only** where Well ID is in **SRBCT - 87 Gene PAM Model**

\*\*\* indicates lists local to this dataset

Subset Label:

**Filter** **Cancel**

## Lab 12. Applying Hierarchical Clustering to the PAM Model

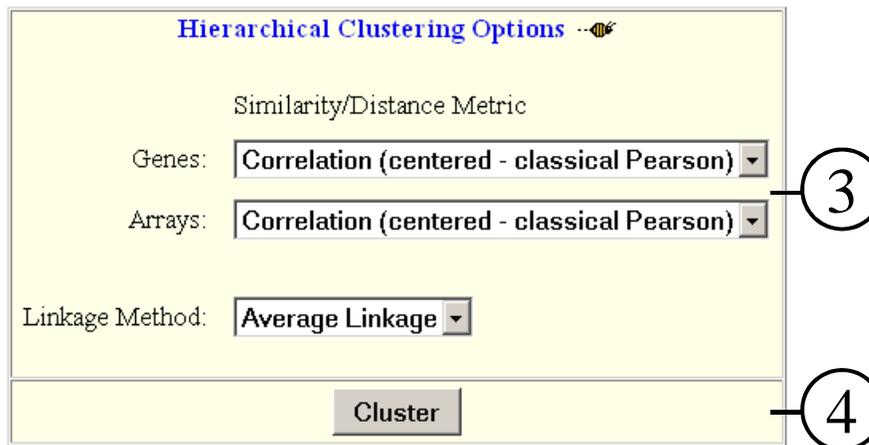


Verify that the current dataset is the right dataset. (87 Gene PAM Model – Complete Dataset )

1. In the Filtering/Grouping/Analysis section, choose the **Clustering: Hierarchical** Tool

2. Click on **Proceed**

A new page will be displayed with options for selecting the Similarity/Distance Metric.



3. Choose **Correlation (centered classical Pearson)** to cluster both **Genes** and **Arrays**.

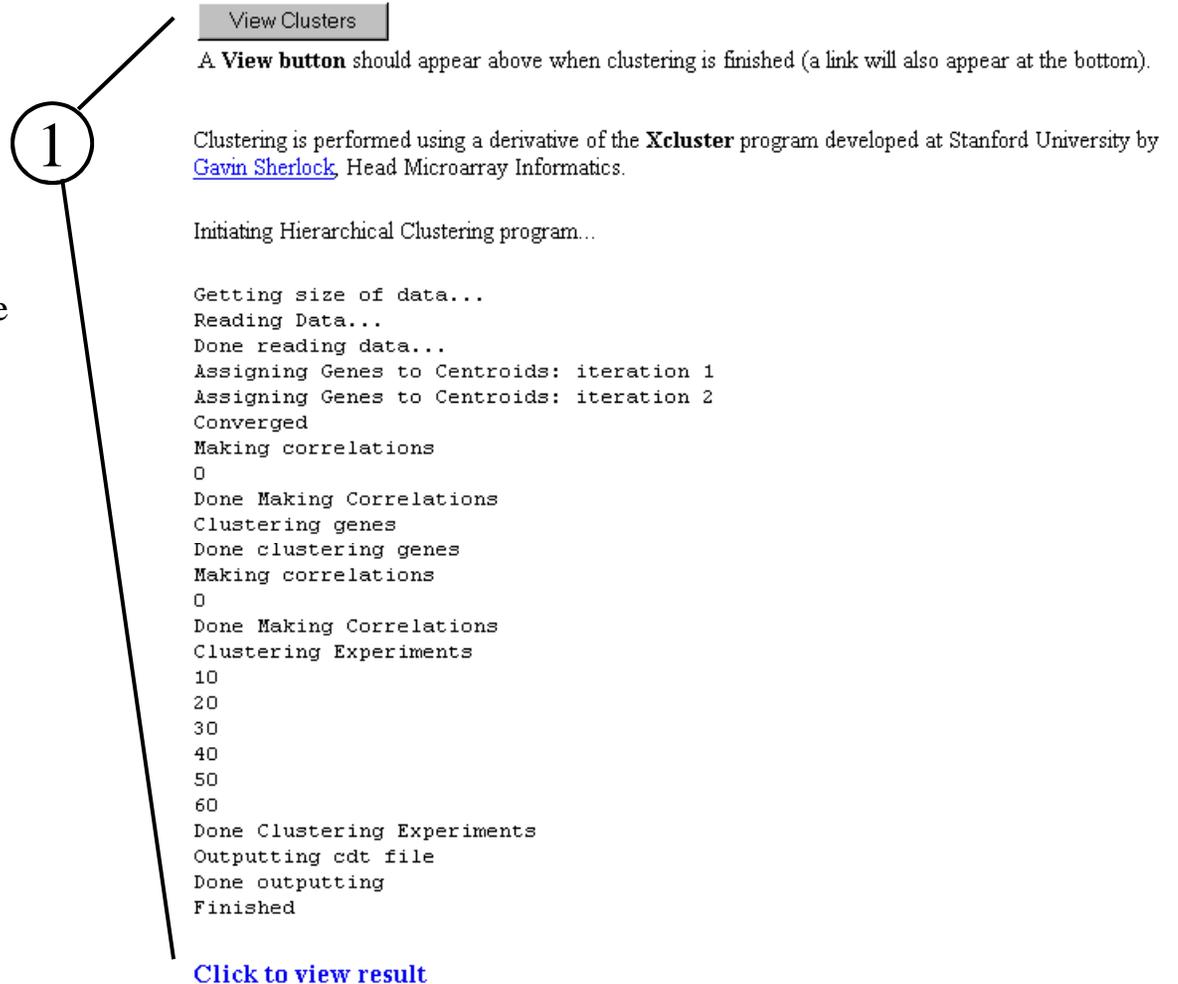
4. Click on **Cluster** button.

## Lab 12. Applying Hierarchical Clustering to the PAM Model

A new page will be displayed for Hierarchical Clustering progress. When the analysis is done, a **View Clusters** button is displayed on top of the page.

1. Click the **View Clusters** button at the top of the page or the **Click to view result** link at the bottom.

A new page will be displayed with a thumbnail image of the clustering results



View Clusters

A **View button** should appear above when clustering is finished (a link will also appear at the bottom).

Clustering is performed using a derivative of the **Xcluster** program developed at Stanford University by [Gavin Sherlock](#), Head Microarray Informatics.

```
Initiating Hierarchical Clustering program...

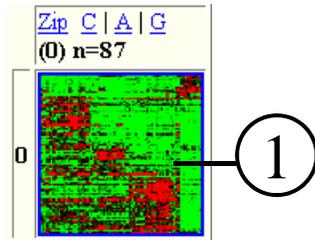
Getting size of data...
Reading Data...
Done reading data...
Assigning Genes to Centroids: iteration 1
Assigning Genes to Centroids: iteration 2
Converged
Making correlations
0
Done Making Correlations
Clustering genes
Done clustering genes
Making correlations
0
Done Making Correlations
Clustering Experiments
10
20
30
40
50
60
Done Clustering Experiments
Outputting cdt file
Done outputting
Finished
```

[Click to view result](#)

## Lab 12. Applying Hierarchical Clustering to the PAM Model

1. Click the thumbnail on the page.

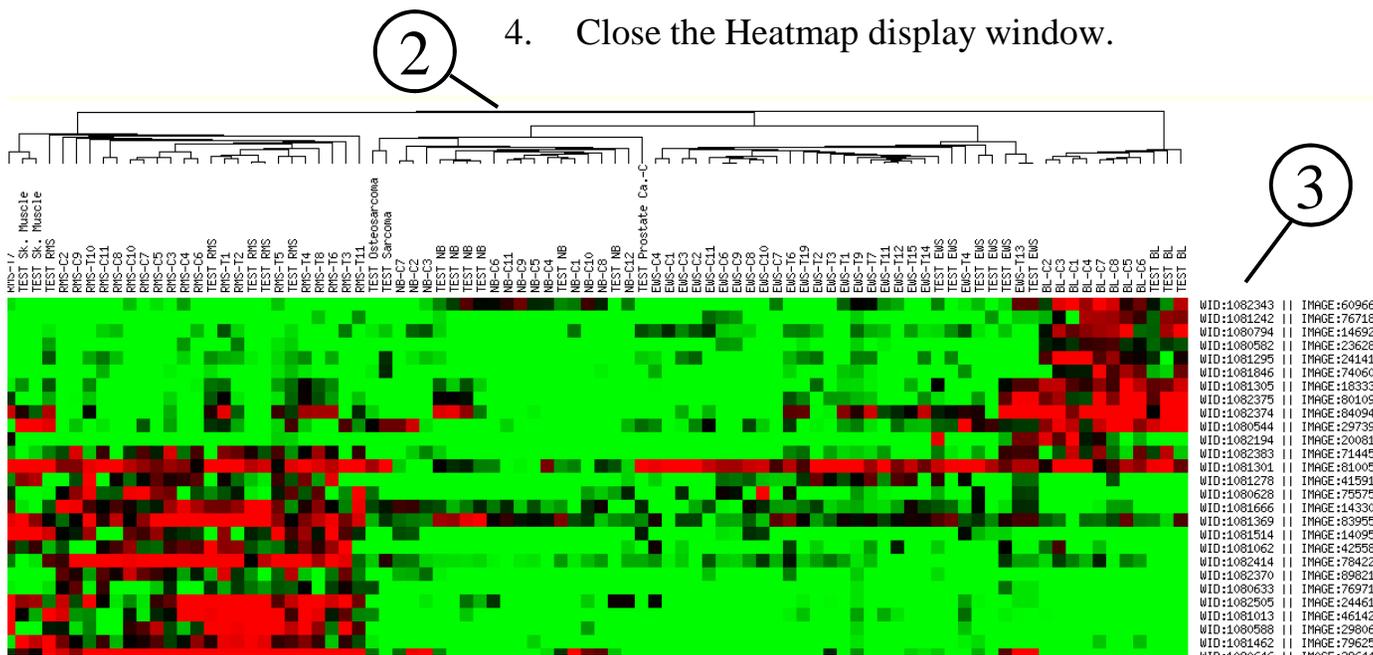
A new browser window will open up to display an enlarged heatmap image and gene tree of the clicked thumbnail image.



2. Check the array tree structure. Check the relationship among all the tumor groups..

3. Click on the gene annotations on the right. A new window will open with a feature report page.

4. Close the Heatmap display window.



### Questions:

1. Review the dendrogram for the samples and identify possible clusters. How does heatmap pattern distinguish the clusters?
2. Review the test arrays not used in the PAM analysis and verify whether they cluster into the right tumor groups.